



American Journal of Pharmacology & Therapeutics

Review Article

The Potential Uses of Insect Biochemicals in Medical Therapeutics -

Cem Turaman *

Free Lance Public Health Specialist (MD), Entomologist (MSc), Postal: İzmir Cad 7/12 Çankaya Ankara, Turkey

***Address for Correspondence:** Cem Turaman, Free Lance Public Health Specialist (MD), Entomologist (MSc), Postal: İzmir Cad 7/12 Çankaya Ankara, Turkey, E-mail: c.turaman19@gmail.com

Submitted: 21 September 2021; **Approved:** 04 October 2021; **Published:** 07 October 2021

Cite this article: Turaman C. The Potential Uses of Insect Biochemicals in Medical Therapeutics. Am J Pharmacol Ther. 2021 Oct 07;6(1): 009-018. doi: 10.37871/ajpt.id26

Copyright: © 2021 Turaman C. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



ABSTRACT

By virtue of research activities, our knowledge on various molecules extracted from the insects that are beneficial to human, is constantly increasing along with the increasing knowledge on the nature of their physiology, morphology and behavior. This article investigates the therapeutic benefits of the insects for humans. The analysed articles chosen out of numerous articles written in this field, shows there are many beneficial secretory or excretory components of the insects having many various effects. Therapeutic molecules that may be beneficial in the treatment of human diseases, probably with much less side effects may be obtained by following this path.

Keywords: Insects; Extracts; Saliva; Venom; Therapeutics

INTRODUCTION

Insect is an arthropod animal with a three sectioned trunk, two pairs of wings and three double pairs of articulated legs with its skeleton outside the body. Insecta is by far the best, adorable species with an incredible biodiversity and ability to adapt to different habitats and environmental conditions. Eventhough there are people who consider the insecta on the lower steps of evolution, many characteristics of their lives show us how developed they are in terms of evolution. One outcome of this evolutionary development is their biodiversity which is much larger than the more evolutionally developed vertebrates. Their high fecundity, their ability to easily adapt to different environments such as high altitudes, poles and deserts, freshwater and seawater; their extraordinary strategies of migration and expansion; their defense mechanisms and living habits of some of their groups are stunning. Apart from their splendid biodiversity and their ability to adapt and colonize new niches and nutritional resources; Insects deserve a special place amongst all living creatures with their strong immune response ability. The physiology, morphology and habits of the Insecta have been used in numerous beneficial ways in the favor of humans and it is certain that this benefit will grow as we get to know Insects better [1]. This article investigates the potential therapeutic benefits of the insects for humans where insects and humans interact. There is a vast literature which investigates the therapeutic value of insects, which proves our growing interest to insects. The examples given below and many other research activities which exceed the limits of an article show that insects have plenty of other secretory or excretory components awaiting to be discovered alongside their anti-viral, anti-fungal and anti-inflammatory effective substances and again effective substances which exist in the insect symbiosis, parasitoid and teratocyte and anti-microbial saliva and venoms. Dr. Tom Eisner from Cronell University states that "it is possible to find numerous new molecules in insects that will occupy the drug industry for years" and defines the research activities carried in this field as "chemical gold rush" [2].

SCOPE

There exist thousands of ramified publications on the subject some of which present classical, some more specified approaches. The aim of this article is to raise pioneering arguments on this wide field, by means of revising of such publications. This pre-analysis aims to highlight the importance of strategic pharmaco-plans through a comprehensive meta-screening of all the research activities on the relative topic with a more systematical approach.

METHODOLOGY

The articles which were considered to be significant by the author were randomly chosen and summarized in order to answer the question of "which new ideas can be derived from this knowledge that would serve for the benefit of humans?"

FINDINGS

Miscellaneous observations

Octopamine and nutrition regulation: The habit of nutrition is a complex behaviour which is organized by means of the nervous system and several chemicals. Octopamine regulates the negative electrical input signals that come from the repressive receptors found in the neural network that covers the crop and oesophagus in the negative direction or inhibits them [3]. These should be post-synaptic facts. The positive signals they send to the central nervous system enable the control of nutrition. These facts which control the crop motility are not completely understood. There are numerous pumps and sphincters which move the nutrition through the digestive system [4].

Magical nuptial feast amongst the angel insects: Zoraptera is a tiny insect group which is closely related to hoppers; they are rare and their adult members might not carry wings. The male *Zorotypus barberi*, a member of this genus, has a cephalic gland in the middle of its head. During courtship the male excretes a liquid from this gland and offers it to the female. The acceptance of this drop by the female relaxes the actions and enables mating [5].

Egg glue of the butterfly: Genus *Opodiphthera* type Saturniidae: Lepidoptera moths stick their eggs on plants with a strong glue. The egg is stuck on the leaves with the help of a rapidly drying glue that deforms the leaves. The glue is seen easily like a kind of membrane which surrounds the mount of the egg. The nature of this glue is unknown and is worth researching. The pupa of the moth also releases the same glue in order to protect the setae of its cremaster. The glue is so strong that it is not possible to detach the silk pillow from the setae [6].

G protein coupled receptors: G Protein Coupled Receptors (GPCR) is a large protein family and is found in almost every eukaryotic organisms. These proteins include numerous receptors which exhibit structural differences and function as mediators that have various functions like sense and feeling regulation, hormone and pheromone production, immune and nervous system functions. These receptors play roles in many human diseases and have become the focus of research activities [7].

Antifreeze proteins of various insects: Thermal hysteresis (the difference between the freezing and melting points) is characteristic of the existence of Antifreeze Proteins (AFP's). AFP's are found in, Chrysomelidae, Silphidae and Coleoptera of the Carabidae family, Neuroptera, Diptera: Tipulidae and Hemiptera *Elasmotethus interstinctus* Pentatomidae, *Limnoporus dissortis* Gerridae type insects. AFP's protect the organism from freezing. Terrestrial arthropods which produce AFP's are protected from freezing. Some

of these AFP producing insects can tolerate freezing under very low temperatures (-40 / -70°C) [8].

Diapause hormone in the flies: Diapause is the spontaneous cease of development and reproduction in some animals caused by the decrease of metabolic activities and dehydration. It is observed in insects, mites, some crustaceans and snails. The phenomenon of the suspension of development is a sign of the approaching negative environmental conditions. Diapause is a behaviour adapted by the insects in order to protect themselves from hard conditions like the winter. Therefore, over-nutrition and increase in the fatty tissue prior to diapause is a 'must'. Insects may enter diapause in any stage of their lives; however diapause during the pupa stage is more common. For the survival of the insect and its species, nutrition-reproduction coordination and the timing of entering and exiting the diapause is crucial. The behaviour of diapause is defined genetically however it might disappear under changing environmental conditions [9]. The levels of chemicals, like sterols decrease in the insects in diapause. In this stage, the sterols enter the ovaries and fill them; this stage does not occur in the females in diapause. One of the hormones regulating the diapause in moths is a 24-aa neuropeptide; which is called as the Diapause Hormone (DH). The cease of DH pupal diapause in the agricultural pest *Helicoverpa/Heliopsis* complex members is accelerated. There are more active DH agonists than the DH's which cease the diapause. One of these agonists prevents the larvae from entering the pupal diapause when given to a larva which is programmed for diapause depending on the environmental conditions [10].

Fly Kuzbanian protein: ADAM's (A Disintegrin and Metalloprotease (Kuzbanian protein)) are a transmembrane protein family and play a central role in transforming the membrane into a soluble form by proteolytic means. Enzymes which have a proteolytic transformation function are believed to take part in the emergence of congenital diseases like the Alzheimer's disease. ADAM's also play a crucial role in the formation of the heart and blood vessels of the vertebrates, thus the mutations in the ADAM genes may be the cause of congenital heart anomalies. The zygotic loss of active Kuzbanian protein form in the fruit fly *Drosophila*, along with the loss of pericardium cells, causes an excessive cardiomyocytes augmentation. Another function of the Kuzbanian may be cardiac-cell morphogenesis [11].

Anaphylaxis treatment with cicada: Anaphylaxis is a chain of reactions which occurs through the release of histamine stemming from the degranulation of mast cells and basophils when an antigen defined as "alien" by the organism holds on to receptors by the IgE's which are immediately released. This may occur in various clinical forms; even though it is rare, it may risk the life of the patient in a state of shock. It is caused by an "alien" drug (penicillin and its derivations, analgesics, iodine-based radiological diagnosis molecules, etc.), pollens or insects bites; rarely less serious cases may occur from food (sesame, eggs, sea products, strawberry, tomatoe, etc.) [12]. *Cryptotympana atrata* Hemiptera is a Cicada species found in Eastern Asia. *Cryptotympana atrata* extract enables a rapid recovery from the anaphylactic shock syndrome. The extract of this insect has proven to prevent histamine release from mice periton mast cells. This extract was used on humans via IV in differing doses and intervals: This extract has proven to prevent histamine release for a long period during continuous, repressive treatments; as well as dramatic conditions like anaphylaxis [13].

Thermolysin of the butterfly: *Galleria mellonella*: Lepidoptera is

known as the greater wax moth. They are considered as pest because their larvae attack honey combs. However, it has a beneficial side because they are used as models for infectious diseases. Even though the insects have no adaptive immune response, they have a very similar one to vertebrates' innate immune response. *G. mellonella*'s immune response is used as model for the diagnosis of Gram-positive and Gram-negative bacteria virulence assessments. Moreover, *G. mellonella* is used in assessing the effect of antimicrobial agents. Thermolysin type metalloproteases like aureolysin, pseudolysin and basillolysin, represent virulence factors of various bacteria and enables innate immune response. Cryptococcosis is a deep mycosis hard to treat which became frequent parallel to the prevalence of HIV infection and is common amongst people whose immune system is depressed. *G. mellonella* is a suitable model organism for *Cryptococcus neoformans* infection. Injecting thermolysin to *Galleria mellonella* larva increases the acquired immunity of the larvae against *Cryptococcus neoformans* infection by means of the small protein fragments which occur due to hemolymph protein proteolysis [14].

Insect-microbe symbiosis

Cockroach: Blattodea have been in a symbiosis with the special bacteroides they carry in their bodies for millions of years. These bacteroides live in a cell called mycetozoid and is transmitted to next generations by the mother cockroach. In return of this comfortable life they lead in the adipose tissue, bacteroides create all the necessary vitamins and amino acids for the insect; this is the secret lying beneath the ability of the cockroaches to transform everything into nutrients. Nitrogen intake and digestion is one of the basic functions of the insects which feed on plants. For example, their ability to produce nitrogen from effluents, offers a crucial resource to insects, in terms of this rare substance, if they acquire the mechanisms which can transform nitrogen [15,16].

Animals stock the excess of uric acid; this would cause problems like kidney diseases and gout in the humans because of their long life span. Uric acid and urea are nitrogen effluents which the animals can't consume as nutrition. However the cockroach which can cooperate with a bacterium is able to use this source in order to produce protein and this means a very distinctive life form; this success depends on the insect-microbe cooperation. *P. americana*, although being a herbivore like many other cockroaches, can become an opportunist carnivore and feed on human wastes. The primary requirement of the cockroach is nitrogen. Other insects eliminate nitrogen as uric acid whereas cockroaches store this substance in their body. Because the cockroach can store nitrogen as uric acid, it can produce nitrogen when needed. According to estimations, for over 140 million years, cockroaches have been in cooperation with a bacterial endosymbiont, *Blattabacterium* that lives in the fatty tissue of their body. This bacterium has two functions: Production of nutrients and keeping the insect from dying by regulating the uric acid level of its body. This bacterium is able to produce all the essential amino acids along with other types of amino acids and various vitamins. It produces nutrients by means of uric acid breakdown products, urea and ammoniac. *Blattabacterium* transforms the waste nitrogen and enables the cockroach to reproduce consuming poor nutrients and benefit from wastes that contain nitrogen; this characteristic may be a speciality which designates the dispersion of cockroach types. *Blattabacterium* is a Gram-negative bacterium which is transmitted vertically. The bacterium does not have uricolytic enzymes but contains urease and glutamate dehydrogenase enzymes, with which it is able to transform uric acid breakdown products into glutamate [17-19].



Beetle: The chemical mediator of *Xyleborus dispar*, Scolytidae: Coleoptera and fungi mutualism is a poly unsaturated peroxide produced by a bacteri. The elective toxicity of this molecule against the insects fungal (ambrosia) antagonist is bacteria-based and indicates a mutually evolved insect-microbe cooperation [20,21].

Wasp: Adult Braconidae: Vespidae is a caterpillar parasitoid and injects its venom and its symbiotic polydnavirus to the *Manduca sexta*: Lepidoptera caterpillar together with its eggs. This venom is not effective on its own; its effect increases in the existence of the polydnavirus. When the eggs hatch in the body cavity of the caterpillar, particular cells produce from their chorion. These cells, called Teratocytes grow into giant cells; the hormones they release decelerate the growth of the host together with the released hormone and the venom. The larva which completes its development in this medium come out of the caterpillar and pupate; later on adult vespids grow out of these pupas. The benefit of the virus from this bargain, which is unable to replicate on its own, is the existence of these genes in the genom of the vespid. The female vespid has particular cells in her ovaries, called calix, and carries virion particles. The viral sequence exists in the male vespid but the virus is not produced in the males. The viral sequence is transmitted to the female during mating, thus the female acquires the ability to replicate virions. Later on, when injected to the host during ovipositioning together with the venom, the Polydnavirus replicates and prevents the growth of the host. The infected host does not go through a metamorphosis but grows into a giant continuing its development and prolongues its life. The injected Polydnavirus also oppresses the immune system of the host caterpillar [22-27].

Butterfly: The effect of *Bacillus thuringiensis* used as a biological insecticide depends on the existence of enteric bacteria in the hemolymph of the insect (*Escherichia coli* and *Enterobacter spp.*). This spore-producing bacteri which can be found anywhere kills the insect larvea by means of insecticide crystal proteins. *Escherichia coli* which is designed for producing *B.thuringiensis* toxin, kills the forest pest *Lymantria dispar* moth caterpillars. *E. coli* and *Enterobacter sp.* can reach up to huge population amounts in the insect hemolymph, contrary to the *B. thuringiensis* which is immediately killed in the insect hemolymph [28].

Parasitoid wasp-host relations

Parasitoid is an organism which lives on or in a host organism and eventually causes its death. Most of the parasitoid insects are included in the Hymenoptera order; approximately 10% of the insect species are parasitoids. Most of them inject their eggs on the host organisms through with their ovipostors. The juveniles that hatch out the egg nurture on the internal organs of the host. The consumed organs are not vital ones at first, thus does not cause the death of the host immediately. Later on, the parasitoid larvae either attacks the vital organs of the host or pupates inside the body of its host; in the first scenario, the host dies immediately and in the second scenario, the host dies while the adult parasitoids hatch out of their pupas. Parasitoids are beneficial in terms of pest control. Potential host insects, on the other hand, have developed various protection mechanisms against this entomophage species. These are effective mechanisms against the predators and parasitoids such as escape, mimicry, repellency and cuticula barrier. These insects also have a second defense line against parasitoids and parasites, called the "immune mechanism". For example, in Africa, *Anopheles gambiae*, the primary vector of human malaria, is able to kill the malaria parasite by enclosing it inside a

melanine-rich capsule in its midgut. Several loci responsible of this encapsulation reaction have been found. This discovery may play a crucial role in malaria control [29-31].

Anaesthesia to the cockroach: Ampulicidae: Vespidae is a solitary and entomophage parasitoid family member of the wasp group mentioned above. One species of this family, *Ampulex compressa* is found in South Asia, Africa and the Pacific islands and shows up during hot months of the year. The mating habit of the female is interesting; she lays its eggs inside the body of a cockroach (*Periplaneta americana* or *Periplaneta australasiae*) which she stings and spaces out two times. The first sting transfuses the venom through the protorasic neural ganglion of the cockroach and hampers it from using its forelegs. When the cockroach becomes immobile, with the second sting, the venom is transfused through the sub-esophageal neural ganglion and the cockroach is no longer able to escape. The active substances of the venom are Gamma Amino-Butyricacid (GABA), taurin and beta-alanine; GABA shows its effect by blocking the neurotransmitter octopamine receptors and thus ceasing the cholinergic transmission. Taurin and beta-alanin, creates an agonist effect prolonging the short-term effect of GABA. Later, the vespid makes use of the effect of the venom and chews the tentacle of the cockroach thus succeeds to immobilize the cockroach without killing it. When she is convinced the effect of the anesthesia is optimal, she drags the cockroach to her nest. There, she lays her eggs inside the abdomen of the cockroach. In order to prevent someone else jerk her precious trophy away, she closes the entrance of her nest with pebbles. When the larvae hatches out of the egg they begin their endoparasitoid lives; they slowly consume the organs of the cockroach keeping it alive long enough and eventually pupate inside the cockroach and finally the adult vespids emerge out of the stomach of the cockroach and begin their new life. The venom shows its basic effect by spoiling the octopaminergic modulation. Octopamine is a neurotransmitter that plays a central role in controlling sudden movements [32-35].

Prevention from the mosquito defense: Parasitoid insects have also developed various methods in order to overcome the potential host defense mechanisms. Encapsulation of the above mentioned alien organisms is a common defense mechanism amongst insects. Co-evolution of the hosts immune response and these protective methods developed against this response, have created a delicate balance between the two. A series of different loci were found amongst anopheles mosquitos which have the ability to create a melanotic encapsulation againts malaria parasites. However, some of the human malaria parasite strains are able to escape from this immune response.

Similar interactions with parasitoids have been proven to exist in *Drosophila melanogaster*: Diptera too, however, parasitoids with high virulence are able to oppress the immune system of this fly. Both of these flies must be encoding the gene products of the encapsulating loci parasitoid recognition and alarm systems in eraly stages. Membrane receptor molecules may be playing a crucial role in the encapsulation process [36-39].

Virus partnership: The polydnaviruses of the endoparasitoid vespid repress the encapsulation reaction of their hosts by means of an unknown molecular mechanism. The beginning of the micro-particle production with the encounter of cell membrane glycoproteins and the virus is a part of the immune response against alien organisms and this is the first step of the cell-mediated encapsulation process. The proteins encoded by the virus may be important means of respressing the host cellular immune response; this method can explain the

repression of the host immune response attacked by a parasitoid [40].

Teratocytes alliance: Various parasitoid insect species, the serosa membrane tears off and produces teratocyte cells when hatching out. Teratocytes are believed to have several functions as immunosuppression, secretion and nutrition in the host-parasitoid relations. The teratocytes growth model reflects whether there is a suitable host or not for the parasitoid. Amongst Braconidae: Hymenoptera species, the teratocyte derives from the serosa membrane of the developing parasitoid embryo. This membrane decomposes into teratocytes while hatching and begins to move through the hemolymph of the host. Teratocytes which develop from trophamnion in most of the parasitoid hymenoptera develops resistance against the defense mechanisms of the host insect. When they are first released from the membrane of the embryo, these cells are little however they grow three thousand times in the circulation of the host. This growth is enabled by nutrition from the host circulation; in return, it represses haematopoiesis in the host hemolymph and protects the juvenil parasitoid from haemocytic activity [41].

Insect antimicrobial peptides

Antimicrobial Peptides (AMP) are multi-functional components of the innate immune system arsenal used both by the prokaryotic and eukaryotic organisms. During recent years, most of these peptides have been isolated from insects. What paved the way for these researches was the observation of the resistance of the insects against bacterial infections. Insects resist the pathogens by means of cellular and especially humoral (innate) immunities. With the identification of the bacteria, an intricate genetic steps system is triggered and finally AMP's are released into the circulation of the hemolymphs. The structures of these peptides are simple and are composed of 20-40 amino acids. As opposed to the differences in terms of their sizes, amino acid sequences and structures, the insect AMP's can be classified under three groups. Peptides rich in proline and glycine are active against gram-negative bacteria; however the defensins kill the Gram-negative bacteria in an elective way, whereas the scorpins are active against both of the bacteria groups. The effect of the insect AMP's are immense; most show its effect by damaging the bacteria membrane or decompose into the membrane structure. AMP's like drosocin, apidaecin and pyrrolicin which show its effect by deactivating the bacterial proteins are exceptions; these stop the growth of the in vivo tumor by means of their cytotoxic specialities. One pyrrolicin analog protected a mouse from experimental *Escherichia coli* infection. Generally, insect AMP's are not cytotoxic and their range of effect is wide however how they effect is not completely understood. The effects may be emerging from peptide-lipid interaction or receptor-mediated recognition. The insect immune system which relies only on innate immunity, work with alternative antimicrobial strategies. One of these strategies is rapid and dense AMP production. However, ability to produce ample and various AMP's is not solely characteristic to insects. Approximately 700 AMP have been recorded in the literature exist also in microorganisms, other animals including mammals and plants [42-44].

Hover flies: Realisation that the antimicrobial peptides existing in Syrphidae and probably other Diptera vastly vary, is promising both in terms of the treatment of dangerous human infections which emerge due to bacteria endotoxins and understanding how closely the molecular evolution of the Diptera genetic immunity and the ecological harmony are related. For example, *Eristalis tenax*,

Syrphidae: Diptera have developed an ability to survive in wet places like cesspits and animal dunghills where microbial pressure is very high. The benefit of living in such places should be protection from predators, opponent species and parasitoids; in return of this evolutionary sacrifice, they managed to invest in immunity against microbial stress, showing that multiple AMP's exist in insects [45].

Black soldier fly: *Hermetia illucens*: Stratiomyidae larval secretion and extract contains substances rich in antimicrobial characteristics which were previously unknown; the characteristics can be crucially effective weapons that can be used against nosocomial infections, especially the Methicilline-Resistant *Staphylococcus aureus* (MRSA) [46]. *H. illucens* larvae, a considerable part of this wide AMP spectrum must be deriving from nutrition with high bacterial loaded diet; meaning, larvae AMP expression depends on the diet. The number of AMP's and the level of AMP expression maximizes when the larvae feed on proteins and sunflower oil. The expansion of the AMP expression repertoire of the larvae should be facilitating its adaption to extraordinary diets thanks to the intestinal microbiota. By force of this ability, it is possible to assume that *H. illucens* is reliable sources for waste control that can be used in organic waste recycling. Using fly maggots in the treatment of wounds that don't heal and multiple-resistant infections is vastly accepted. It is possible to produce antibacterial substances against Gram-positive *Staphylococcus aureus*, MRSA and Gram-negative *Pseudomonas aeruginosa* from *Hermetia illucens* larvae. That is to say, the larvae extract exhibits a wide antibacterial spectrum, especially against MRSA, *H. illucens* larvae secretions may be promising instead of antibiotics [47,48].

Insect saliva

Hematophagy is a convergent behavior in evolution; it developed independently in worms, arthropods and mammals. Each one of them has developed different mechanisms in order to inactivate host defense, thus these mechanisms are specific to genus and the family and show an immense abundance. The saliva of the blood sucking insects contains hundreds of polypeptides. One of these nutrition-dependent adaptations is the equipment of these insect saliva molecules which derive their hosts from haemostasis, inflammation and immune response weapons. Insect saliva contains hundreds of polypeptides which are mostly specific to their own genus or family [49,50].

Mosquito: The blood sucking arthropod saliva contains a clotting inhibitor, an antiplatelet agent and a vasodilator substance as a rule. Mosquito saliva, (as in tick and phlebotominae) apart from its ability to retard host vessel collaps, to prevent haemostatic and immune responses, also decreases the early expression of interferon α/β , thus enables reproduction of viruses like the West Nile Virus within its host. The components of the saliva which prevent clotting can be friendlier and effective agents compared to clotting preventing and vasodilator agents that are currently being used in medicine. Mosquito saliva can also be beneficial in controlling the bacteria that reproduces in sugary environments because it contains enzymes that help consumption of the glyucose in the blood of vertebrates. Mosquito saliva composition is quite simple, contains less than 20 essential proteins. Despite the well-rounded knowledge on these molecules, what more than half them serves is still unknown. For example, producing coagulation-preventing and capillary expanding agents can be very helpful in the control of cardiovascular diseases because they are consumer-friendlier compared to the drugs in use [51-54].

Assasin bug: Reduviid predators *Rhynocoris marginatus* (Fab.)

and *Catamirus brevipennis* (Servile) use their venoms to paralyze their prey. Both the venoms are proteins with low molecular weight. These two Reduviid venoms have been proven to have antibacterial activities against seven gram-negative and four gram-positive bacteria. The venoms of these two insects exhibit a strong antibacterial activity by preventing the bacteria to nurture and also preventing the aggregation of haemocytes. The *R. marginatus* venom has shown antibacterial activity against *Escherichia coli*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Salmonella typhimurium*, *Streptococcus pyogenes*; and the *C. brevipennis* venom has shown antibacterial activity against *Escherichia coli*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Proteus vulgaris*, *Salmonella typhimurium*, *Bacillus subtilis*, *Staphylococcus aureus* and *Bacillus sphaericus* [55,56].

Venomous insect secretions

Arctiidae butterfly aposematism: Most of the animal species, consume chemicals which taste bad or poisonous from the plants they live on, apart from the nutritious substances they consume. Some of them produce defensive substances from these chemicals. The most important of such chemicals are cardiac glycosides, pyrrolizidine alkaloids, pyrazines and histamins. Aposematism is the term used for indicating that a living creature is dangerously poisonous or has a very bad taste; these have very bright colours. A good example for this is the Cinnabar: Arctiidae Lepidoptera moth larvae. There are horizontal orange/black stripes on the body of this larva. *Jacobaea vulgaris* plants on which these caterpillars feed is rich in alkaloids. The caterpillar that feeds on the leaves of this plant accumulates this poison and preserves the alkaloid until it becomes adult. The aposematic colouring of the caterpillar indicates it is poisonous and warns the predators. Arctiidae: Lepidoptera larvae and the adult have a complex allelochemical sequestration strategy. Arctiid uses two mutually independent defensive signs in courtship; these are secondary chemicals and the ultrasound. Pyrrolizidine alkaloids are the defensive tool of the larvae, however they have become the courtship pheromone of the adult male in most of the lineages. The ultrasound is developed as a means of protection from the bats and is also being used in courtship games [57-61].

Lonomia butterfly: Hemileucinae: Saturniidae caterpillar lives as a community on a host tree, they have stinging bristles and those of *Lonomia* contain a poison which can be fatal for humans. *Lonomia* genus caterpillar venom is one of the most fatal animal venoms and its LD₅₀ value has not been measured so far. A typical intoxication with the caterpillar of this moth takes place if the victim leans against the host tree. Because the tree hosts numerous caterpillars, the clinical picture can be dramatic; it results in disseminated intravascular coagulation and consumption coagulopathy, hemorrhagic syndrome; massive internal hemorrhage, renal failure and hemolysis are the components of this clinical picture. This syndrome is called Lonomiasis and has been reported in 1967 for the first time. *Lonomia oblique* caterpillar venom exists in the sacs at the bottom of the spikes on its body. These spikes which are empty inside like a pipe, when stung to a vertebrate, transfuse the venom to the victim. The effect of this venom emerges as prevention of blood clotting. In order to be effected the victim should be stung 20-100 times; because the caterpillars live as a community it shouldn't be hard to be stung so many times. The fatality rate has been reported as 1.7-2.5%. An anti-venom is being produced in Brazil [62-65].

Pederin: Molecules which can potentially be used as drugs, tested on numerous marine and terrestrial invertebrates, are probably the metabolites of bacterial symbionts of these organisms. For example, anti-tumor polycetides pederin family isolated from coleoptera and sponges, indicates an evolutionary relation between the two species. Pederin is a strong DNA inhibitor which exists in the hemolymph circulation of some Paederus, Paederinae: Staphylinidae species in all stages of their lives. This molecule is produced by the endosymbionts of some adult females and is transferred to their eggs; thus is transfused to the larvae and the pupa. The male insect can only procure this molecule by consuming the eggs. Pederin is an effective defense mechanism against predator spiders, however despite the fact that it is known as one of the most powerful animal toxins, its insecticide effect has not been proven so far. It causes dermatitis in humans and causes heavy tissue damage if it contacts the eye. If these strong effects can be repressed, it might be possible to use it in healing chronic wounds and ceasing tumoral growth. The examples of this species elevated under laboratory conditions are aposymbionts; they can not biosynthesize venom molecules because they don't have endosymbionts [66-71].

Cantharidin: Another Coleoptera genus, *Epicauta*, exists in trefoils; this is highly toxic for the horses that feed on trefoils and may cause the death of the horse. One other coleopter species *Lytta vesicatoria*, have been used in traditional medicine exercises and poisoning cases have been reported. The active agent of this insect venom is cantharidin and when it contacts the skin it causes dermatitis. We know that 449 species including the Oedemeridae: Coleoptera family, produce cantharidin. The lethal dose of cantharidin for the humans is as small as 0.03g. Symptoms of intoxication are oversalivation, headache, vomiting and bloody diarrhea; central nervous system symptoms can also be observed but death is rare. It has been used in traditional medicine in the treatment of rheumatism, pneumonia, inflammation and gout. It is also being used as a love potion and most of the intoxication cases emerge as a result of abusive usage as such. Cantharidin is being used in modern medicine in the treatment of molluscum contagiosum [27,72,73].

Lucibufagins: Lampyridae are known as unsavory and sometimes poisonous by the vertebrates and are not consumed. A steroid pyrone group known as lucibufagins is responsible for this protection and its effect is similar to the cardiotonic bufadienolides existent in poisonous salientians. Bufadienolides are a steroid hormone group, they are found in blood and are eliminated through urine. Their effect emerges by means of adenosine triphosphatase sodium-potassium pump inhibition. This effect causes an increase of the sodium elimination, vasoconstriction and hypertension; they work as cardiac inotropes. Bufadienolides are responsible for hypertension related to volume expansion; they hinder cell proliferation and inhibit cell maturation. Resibufogenin, an antagonist of one of the most analysed bufadienolides marinobufagenin, can be beneficial in the treatment of diseases where high levels of marinobufagenin plays a role in their pathophysiology. For example, preeclampsia is a condition with heavy fetomaternal complications and with no definitive treatment. Marinobufagenin should be playing a role in preeclampsia etiology and can be easily used among pregnant women to screen preeclampsia signs in the urine [74-76].

Mastoparan: Vespidae Hymenoptera is a large, varied, cosmopolitan and spinule wasp family with nearly 5000 different species; almost all the social wasps are included in this group. Mastoparan is a peptide



extracted from vespid venom and is probably the active agent of the vespid venom. It shows its effect by directly increasing the GTPase activity and the G proteins. The effect of the venom depends on the type of the cell, however exocytosis is the main phenomenon. Mastoparan causes histamine release from mast cells; it causes serotonin release from thrombocytes and catecholamine release from chromaffin cells, prolactin release from anterior hypophysis [77].

CONCLUSION AND SUGGESTIONS

ADAM protein which is existent in vertebrates and plays a central role in the formation of heart, also plays an important role in the proteolytic transformation of the membrane binding proteins in insects. Proteolytic transformation enzymes are believed to play a crucial role in congenital diseases. Kuzabanin protein can be used in the treatment of Alzheimer and cardiac birth defects.

Octopamine is believed to regulate nutrition in insects. Octopamine is used as a sympathomimetic in emergency medicine. It might be used for regulating the digestive system activities in lower dosages.

AFP's (Anti Freeze Proteins) are shown to exist in 75 different insect species's (Chrysomelidae, Silphidae, Carabidae, Neuroptera, Tipulidae, Tingidae, Pentatomidae, Gerridae) hemolymph in the Arctic Region and their effective mechanisms are still subject to argumentations. In exposure injuries, purified AFP can be a life saver. Scientists who risk to be exposed to cold (not military brigades!) can be premedicated with AFP.

When the sterol levels of Musca: Diptera in diapause decrease, the ovarial development ceases. It might be an opportunity to decide about contraception and time to conceive? Several agonists have been developed inspired from DH structure and from these, DH antagonists were produced which aims to control the pest species that enter diapause during winter. By this means, entering the diapause in these pests are prevented. Preventing the pest species from surviving the winter can be an effective pest control strategy. In manic depressive psychosis, respectively DH agonists (manic period) and antagonists (depressive period); also in sleeping disorders, attention deficit disorders and hyperactivity disorders, DH agonists and antagonists can be a solution.

If the behaviour observed on female zoraptera are dependent on the psychoactive effect of the male cephalic gland components, it may be beneficial to analyse how this effect functions and to evaluate it on humans, and to use it for anaesthesia and the control of psychiatric disorders.

Study of the interaction of food-borne bacterial infections which cause dangerous intestinal diseases in humans with *Bacillus thuringiensis*, can be helpful in the control of *E. coli* toxic shock syndrome.

The effect mechanism of *Cryptotympana atrata*: Hemiptera extract, suggests that this extract may be used in the treatment of auto-immune diseases. Presuming that approximately 1/3 of the humans face an auto-immune disease in their lives, the importance of a simple and inexpensive treatment becomes evident. Treatment with *Cryptotympana atrata* extract, awaits clinical research studies.

Usage of Mastoparan can be used in the treatment of diseases caused by the lack of histamine release, in increasing the effect of mental functions and control of sleep.

The venom of vespid contains GABA, taurine and beta-alanine which accomplish its effect. These substances, when injected at the right place, block the motor nervous action potential temporarily. This blockage emerges through repression of cholinergic transmission by increasing the chloride conductivity in the synapses. It is worth the try to use it in local anesthesia, sympathectomy practices (Buerger disease, electric injuries, viperidae bites) and pain control in some cancers.

In AIDS patients, along with antimicrobics implementations, the success of the treatment depends on strengthening the immune system and preservation of T4/T8 balance is crucial. Thermolysin can be an option in some cases.

Insect egg glues can be useful in curing operation wounds and other clean wounds.

Cockroaches benefit from the vitamins produced by bacteria, in return to supplying the bacteria a living space. In Coleoptera, the polyunsaturated peroxide produced by bacteria has a selective toxicity against fungal antagonists in insects. A way to producing natural vitamins and antimicrobial agents.

Antibiotic-producing microbe-insect mutualism can pave the way for producing antimicrobial components that can be used for treatment of infectious diseases.

The ability to kill Plasmodium by producing a melanin-rich capsule in the midgut of the *Anopheles gambiae* can present an important opportunity in malaria control.

Insect parasites and parasitoids, prevent the immune response of their host insect by means of an unknown effect they create in membrane receptor molecules, thus prevent encapsulation response of the host. Preventing the encapsulation response of the host insect emerges by the repression of the cellular immune system with the effect of the polydnavirus-transformed proteins. The mechanism of Ichnemonid that prevents the cellular immune response by use of polydnavirus can be used in preventing tissue rejection in organ transplantations.

Braconid, by injecting together the venom and symbiotic virus to its host, produces hormones that prevent the growth of the host; teratocytes grow and work in cooperation with the virus in this process. What causes the reproduction of the virus is the Braconid genome. It can be a friendly immune response repressor in organ transplantations.

Teratocyte cells have several mediator functions like immunosuppression on parasitoid species. In Braconid species, they wrap the developing parasitoid embryo and during ecdyses, it transfuses in the host haemocoel by resolving into individual cells. The basic role of the giant teratocytes which grow from the trophamnion in parasitoid hymenoptera is to resist the host defense mechanisms. These cells which are small while they resolve inside the embryo membrane, then transfuses in the host haemocoel and grow by means of the nutrients existent in the micro-environment. The host hematopoiesis is repressed depending on the existence of teratocytes and the effectiveness of the hemocyte response is decreased. Apart from the diseases related to humoral immunity, the ones that are characterized with the over-increase of hematopoiesis, myeloproliferative diseases like polycythemia, hematological malignancies like lymphomas, hemoglobinopathies, hemophilia, coagulopathies, AIDS (which indirectly effect the hematopoietic



system even though it is related to cellular immunity), autoimmune diseases can be controlled with teratocytes.

It may be possible to create user-friendly alternatives to antimicrobials by selecting and evaluating the molecules extracted from insect AMP's and expanding their scale of use. AMP existence in humans should also be investigated. Understanding their effects on preventing and treating infections can be a pioneering discovery.

What conclusions can be drawn from the fact that the mosquito saliva prevents interferon response? It can be an opportunity in preventing immune response in organ transplantation. It is certain that it plays a crucial role in preventing and treating diseases related to thromboembolism; taking into consideration the extensity of deaths related to cardiovascular diseases, the importance of this area is not even questionable. It can also be an important agent in controlling the blood sugar. It can be beneficial in controlling the sugar-consuming ferment activity in wine and beer production.

Reduviidae saliva can be an effective biological agent in treatment of local and systemic infections and pest control. Its value as an anaesthetic should also be analysed.

Cinnabar: Arctiidae moth larvae takes pyrrolizidine alkaloides from the plants they consume and use them in defense; this substance turns into pheromones used by the adult male at the end of the metamorphose. It is used extensively for inflammatory diseases in Chinese medicine. Its antimicrobial effect should be investigated.

Numerous studies have been made in order to understand the medical value of *Lonomia oblique* caterpillars' venom. Especially, the anticoagulant and anti-apoptotic characteristic of the "lopap" (*L. obliqua* prothrombin activator protease) component of this venom is worth the attention. The most dangerous species of the family is *Lonomia obliqua* (Walker), therefore the pharmacological research are focused on this species. It may be useful in preventing coronary thrombosis by means of its anticoagulant effect.

Pederin is a strong toxin. It may have a strong effect on cancers and chronic wounds. It increases the effect of the used antibiotics. It may also be useful in antitoxin and anatoxin production. How come the toxin circulating in the Paederinae: Staphylinidae hemolymph, without harming the insect? A possible anti-pederine substance may be as important as the pederine itself. This molecule can be life saving in antibiotic resistant Gram-negative sepsis. Understanding the antimitotic role of pederine can be useful in developing a user-friendly drug in cancer treatments.

The highly toxic and caustic effect of cantharidin can emerge through consummation of and contact with the Coleopter. It effects bristle growth and was used in traditional medicine. Its toxic effect can be useful in producing antitoxin and anatoxin. Its antimitotic activity should be investigated. Its probabile aphrodisiac effect can be a huge commercial effect. Its sympathomimetic activity can be beneficial in some intoxications.

The structure of lucibufagins are similar to those of cardiacglycosides and has a cardiotonic effect. It may be playing a role in preeclampsia etiopatology. Its cardiac inotrop effect can be useful in shock treatment. It can be used a scanning test for preeclampsia. If its antitoxin can be produced, it can be a user-friendly drug in hypertension and hyperemesis gravidarum treatment.

REFERENCES

- Borror DJ, DeLong DM, Johnson NF. An introduction to the study of insects. 6th ed. Philadelphia: Saunders College Publishing; 1989. p. 875. <https://tinyurl.com/x99mwz3y>
- Eisner, T, For Love of Insects. Harvard University Press. 2003 ISBN 0-674-01181-3. <https://tinyurl.com/4bhntb4d>
- Long TF and Murdock LL. Stimulation of blowfly feeding behavior by octopaminergic drugs. Proc. Natl. Acad. Sci. 1983;80:4159-4163. doi: 10.1073/pnas.80.13.4159
- Brookhart GL, Edgecomb RS, Murdock LL. Amphetamine and reserpine deplete brain biogenic amines and alter blow fly feeding behavior. Jour. Neurochemistry. 1987;48:1307-1315. doi: 10.1111/j.1471-4159.1987.tb05662.x
- Choe JC. The evolution of mating systems in the Zoraptera: Mating variations and sexual conflicts. In: CHOE JC & CRESPI BJ, editors. The evolution of mating systems in insects and Arachnids. Cambridge University Press: Cambridge, 1997. p.130-145.
- Li D, Huson MG, Graham LD. Proteinaceous adhesive secretions from insects, and in particular the egg attachment glue of Opodiphthera sp. moths. Arch Insect Biochem Physiol. 2008 Oct;69(2):85-105. doi: 10.1002/arch.20267. PMID: 18780346.
- Filmore D. It's a GPCR world. Modern drug discovery 2004;7:24-28. <https://tinyurl.com/8nk3443a>
- Duman JG, Bennett V, Sformo T, Hochstrasser R, Barnes BM. Antifreeze proteins in Alaskan insects and spiders. J Insect Physiol. 2004 Apr;50(4):259-66. doi: 10.1016/j.jinsphys.2003.12.003. PMID: 15081818.
- Stoffolano J. The synchronization of the life cycle of diapausing face flies, *Musca autumnalis*, and of the nematode *Heterotylenchus autumnalis*. J Invert Pathol. 1967;9:395-397. doi: 10.1016/0022-2011(67)90076-6
- Zhang Q, Nachman RJ, Kaczmarek K, Zabrocki J, Denlinger DL. Disruption of insect diapause using agonists and an antagonist of diapause hormone. Proc Natl Acad Sci U S A. 2011 Oct 11;108(41):16922-6. doi: 10.1073/pnas.1113863108. Epub 2011 Sep 22. PMID: 21940497; PMCID: PMC3193214.
- Albrecht S, Wang S, Holz A, Bergter A, Paululat A. The ADAM metalloprotease Kuzbanian is crucial for proper heart formation in *Drosophila melanogaster*. Mech Dev. 2006 May;123(5):372-87. doi: 10.1016/j.mod.2006.03.005. Epub 2006 Mar 27. PMID: 16713197.
- White JM, Rumbold GR. Behavioural effects of histamine and its antagonists: a review. Psychopharmacology (Berl). 1988;95(1):1-14. doi: 10.1007/BF00212757. PMID: 3133686.
- Shin TY, Park JH, Kim HM. Effect of *Cryptotympana atrata* extract on compound 48/80-induced anaphylactic reactions. J Ethnopharmacol. 1999 Sep;66(3):319-25. doi: 10.1016/s0378-8741(98)00223-2. PMID: 10473179.
- Altincicek B, Linder M, Linder D, Preissner KT, Vilcinskis A. Microbial metalloproteinases mediate sensing of invading pathogens and activate innate immune responses in the lepidopteran model host *Galleria mellonella*. Infect Immun. 2007 Jan;75(1):175-83. doi: 10.1128/IAI.01385-06. Epub 2006 Oct 30. PMID: 17074843; PMCID: PMC1828416.
- Dybas C. With help from a bacterium, cockroaches develop way to store excess uric acid. NSF 2019;(703):292-7734. <https://tinyurl.com/yb64vd84>
- Moran NA, Plague GR, Sandström JP, Wilcox JL. A genomic perspective on nutrient provisioning by bacterial symbionts of insects. Proc Natl Acad Sci U S A. 2003 Nov 25;100 Suppl 2(Suppl 2):14543-8. doi: 10.1073/pnas.2135345100. Epub 2003 Oct 3. PMID: 14527994; PMCID: PMC304116.
- Sabree ZL, Kambhampati S, Moran NA. Nitrogen recycling and nutritional provisioning by *Blattabacterium*, the cockroach endosymbiont. Proc Natl Acad Sci U S A. 2009 Nov 17;106(46):19521-6. doi: 10.1073/pnas.0907504106. Epub 2009 Oct 30. PMID: 19880743; PMCID: PMC2780778.
- Ayayee PA, Larsen T, Sabree Z. Symbiotic essential amino acids provisioning in the American cockroach, *Periplaneta americana* (Linnaeus) under various dietary conditions. PeerJ. 2016 May 18;4:e2046. doi: 10.7717/peerj.2046. PMID: 27231663; PMCID: PMC4878363.
- Ayayee PA, Ondrejch A, Keeney G, Muñoz-Garcia A. The role of gut microbiota in the regulation of standard metabolic rate in female *Periplaneta*



- americana. PeerJ. 2018 May 24;6:e4717. doi: 10.7717/peerj.4717. PMID: 29844953; PMCID: PMC5971104.
20. Malloch D, Blackwell M. Dispersal biology of ophiostomatoid fungi. In: Ceratocystis and Ophiostoma: Taxonomy, Ecology and Pathology. Wingfield MJ, Seifert KA, Webber JF, editors. APS, St. Paul; 1993. p. 195-206.
21. Francke-Grossmann H. Ectosymbiosis in wood inhabiting insects. In: Symbiosis. Henry M, editor. New York: Academic Press; 1967. p. 141-205.
22. Beckage NE, Tan FF, Schleifer KW, Lane RD, Cherubin LL. Characterization and biological effects of cotesia congregata polydnavirus on host larvae of the tobacco hornworm, manduca sexta. Archives of Insect Biochemistry and Physiology 1994;26:165-195. doi: 10.1002/arch.940260209
23. de Buron I, Beckage NE. Characterization of a polydnavirus (PDV) and virus-like filamentous particle (VLFP) in the braconid wasp Cotesia congregata (Hymenoptera: Braconidae). Journal of Invertebrate Pathology. 1992;59:315-327. doi: 10.1016/0022-2011(92)90139-U
24. Wyler T, Lanzrein B. Ovary development and polydnavirus morphogenesis in the parasitic wasp Chelonus inanitus. II. Ultrastructural analysis of calyx cell development, virion formation and release. J Gen Virol. 2003 May;84(Pt 5):1151-1163. doi: 10.1099/vir.0.18830-0. PMID: 12692280.
25. Marti D, Grossniklaus-Bürgin C, Wyder S, Wyler T, Lanzrein B. Ovary development and polydnavirus morphogenesis in the parasitic wasp Chelonus inanitus. I. Ovary morphogenesis, amplification of viral DNA and ecdysteroid titres. J Gen Virol. 2003 May;84(Pt 5):1141-1150. doi: 10.1099/vir.0.18832-0. PMID: 12692279.
26. Lavine MD, Beckage NE: Temporal pattern of parasitism-induced immunosuppression in Manduca sexta larvae parasitized by Cotesia congregata. Journal of Insect Physiology. 1996;42:41-51. doi: 10.1016/0022-1910(95)00081-X
27. Zitnan D, Kingan TG, Kramer SJ, Beckage NE. Accumulation of neuropeptides in the cerebral neurosecretory system of Manduca sexta larvae parasitized by the braconid wasp Cotesia congregata. J Comp Neurol. 1995 May 22;356(1):83-100. doi: 10.1002/cne.903560106. PMID: 7629311.
28. Broderick NA, Raffa KF, Handelsman J. Midgut bacteria required for Bacillus thuringiensis insecticidal activity. Proc Natl Acad Sci U S A. 2006 Oct 10;103(41):15196-9. doi: 10.1073/pnas.0604865103. Epub 2006 Sep 27. PMID: 17005725; PMCID: PMC1622799.
29. Vinson SB. How parasitoids deal with the immune system of their host: An overview. Arch. Insect Biochem. Physiol. 1990;13:3-27. doi: 10.1002/arch.940130103
30. Zheng L. Genetic basis of encapsulation response in Anopheles gambiae. Parasitologia. 1999 Sep;41(1-3):181-4. PMID: 10697853.
31. Asgari S, Schmidt O, Theopold U. A polydnavirus-encoded protein of an endoparasitoid wasp is an immune suppressor. J Gen Virol. 1997 Nov;78 (Pt 11):3061-70. doi: 10.1099/0022-1317-78-11-3061. PMID: 9367394.
32. Williams FX. *Ampulex compressa* (Fabr.), a cockroach-hunting wasp introduced from New Caledonia into Hawaii. Proc. Hawaiian Entomological Society. 1942;11:221-233. <https://tinyurl.com/4c83pdwr>
33. Haspel G, Rosenberg LA, Libersat F. Direct injection of venom by a predatory wasp into cockroach brain. J Neurobiol. 2003 Sep 5;56(3):287-92. doi: 10.1002/neu.10238. PMID: 12884267.
34. Moore EL, Haspel G, Libersat F, Adams ME. Parasitoid wasp sting: a cocktail of GABA, taurine, and beta-alanine opens chloride channels for central synaptic block and transient paralysis of a cockroach host. J Neurobiol. 2006 Jul;66(8):811-20. doi: 10.1002/neu.20254. PMID: 16673394.
35. Gal R, Rosenberg LA, Libersat F. Parasitoid wasp uses a venom cocktail injected into the brain to manipulate the behavior and metabolism of its cockroach prey. Arch Insect Biochem Physiol. 2005 Dec;60(4):198-208. doi: 10.1002/arch.20092. PMID: 16304619.
36. Adamo SA. The specificity of behavioral fever in the cricket Acheta domesticus. J Parasitol. 1998 Jun;84(3):529-33. PMID: 9645851.
37. Banford S, Thomas MB, Langeward J. Behavioral fever in the Senegalese grasshopper, Oedaleus senegalensis, and its implication for biological control using pathogens. Ecol. Entomol. 1998;23:9-14. <https://tinyurl.com/8hns3js3>
38. Bunday S, Raymond S, Dean P, Roberts SK, Dillon RJ, Charnley AK. Eicosanoid involvement in the regulation of behavioral fever in the desert locust, Schistocerca gregaria. Arch Insect Biochem Physiol. 2003 Apr;52(4):183-92. doi: 10.1002/arch.10081. PMID: 12655606.
39. Zheng L. Genetic basis of encapsulation response in Anopheles gambiae. Parasitologia. 1999 Sep;41(1-3):181-4. PMID: 10697853.
40. Asgari S, Schmidt O, Theopold U. A polydnavirus-encoded protein of an endoparasitoid wasp is an immune suppressor. J Gen Virol. 1997 Nov;78 (Pt 11):3061-70. doi: 10.1099/0022-1317-78-11-3061. PMID: 9367394.
41. Firlej A, Lucas É, Coderre D, Boivin G. Teratocytes growth pattern reflects host suitability in a host-parasitoid assemblage. Physiological Entomology. 2007;32:181-187. doi: 10.1111/j.1365-3032.2006.00548.x
42. Otvos L Jr. Antibacterial peptides isolated from insects. J Pept Sci. 2000 Oct;6(10):497-511. doi: 10.1002/1099-1387(200010)6:10<497::AID-PSC277>3.0.CO;2-W. PMID: 11071264.
43. Bulet P, Hetru C, Dimarcq JL, Hoffmann D. Antimicrobial peptides in insects; structure and function. Dev Comp Immunol. 1999 Jun-Jul;23(4-5):329-44. doi: 10.1016/s0145-305x(99)00015-4. PMID: 10426426.
44. Bulet P, Stöcklin R. Insect antimicrobial peptides: structures, properties and gene regulation. Protein Pept Lett. 2005 Jan;12(1):3-11. doi: 10.2174/0929866053406011. PMID: 15638797.
45. Altincicek B, Vilcinskas A. Analysis of the immune-inducible transcriptome from microbial stress resistant, rat-tailed maggots of the drone fly Eristalis tenax. BMC Genomics. 2007 Sep 17;8:326. doi: 10.1186/1471-2164-8-326. PMID: 17875201; PMCID: PMC2039750.
46. Duman JG, Bennett V, Sformo T, Hochstrasser R, Barnes BM. Antifreeze proteins in Alaskan insects and spiders. J Insect Physiol. 2004 Apr;50(4):259-66. doi: 10.1016/j.jinsphys.2003.12.003. PMID: 15081818.
47. Park S, Chang BS, Yoe SM. Antimicrobial activity of *H. illucens*. Entomological Research. 2014;44:58-64.
48. Vogel H, Müller A, Heckel DG, Gutzeit H, Vilcinskas A. Nutritional immunology: Diversification and diet-dependent expression of antimicrobial peptides in the black soldier fly Hermetia illucens. Dev Comp Immunol. 2018 Jan;78:141-148. doi: 10.1016/j.dci.2017.09.008. Epub 2017 Sep 29. PMID: 28966127.
49. Alves-Silva J, Ribeiro JM, Van Den Abbeele J, Attardo G, Hao Z, Haines LR, Soares MB, Berriman M, Aksoy S, Lehane MJ. An insight into the sialome of Glossina morsitans morsitans. BMC Genomics. 2010 Mar 30;11:213. doi: 10.1186/1471-2164-11-213. PMID: 20353571; PMCID: PMC2853526.
50. Ribeiro JM, Francischetti IM: Role of arthropod saliva in blood feeding: sialome and post-sialome perspectives". Annual Review of Entomology 2003;48:73-88 doi: 10.1146/annurev.ento.48.060402.102812
51. Ribeiro JM, Francischetti IM. Role of arthropod saliva in blood feeding: sialome and post-sialome perspectives. Annu Rev Entomol. 2003;48:73-88. doi: 10.1146/annurev.ento.48.060402.102812. Epub 2002 Jun 4. PMID: 12194906.
52. Grossman GL, James AA. The salivary glands of the vector mosquito, Aedes aegypti, express a novel member of the amylase gene family. Insect Mol Biol. 1993;1(4):223-32. doi: 10.1111/j.1365-2583.1993.tb00095.x. PMID: 7505701.
53. Rossignol PA, Lueders AM. Bacteriolytic factor in the salivary glands of Aedes aegypti. Comp Biochem Physiol B. 1986;83(4):819-22. doi: 10.1016/0305-0491(86)90153-7. PMID: 3519067.
54. Schneider BS, Higgs S. The enhancement of arbovirus transmission and disease by mosquito saliva is associated with modulation of the host immune response. Trans R Soc Trop Med Hyg. 2008 May;102(5):400-8. doi: 10.1016/j.trstmh.2008.01.024. Epub 2008 Mar 14. PMID: 18342898; PMCID: PMC2561286.
55. Valenzuela JG, Pham VM, Garfield MK, Francischetti IM, Ribeiro JM. Toward a description of the sialome of the adult female mosquito Aedes aegypti. Insect Biochem Mol Biol. 2002 Sep;32(9):1101-22. doi: 10.1016/s0965-1748(02)00047-4. PMID: 12213246.
56. Sahayaraj K, Muthukumar S. Zootoxic effects of reduviid Rhynocoris marginatus (Fab.) (Hemiptera: Reduviidae) venomous saliva on Spodoptera litura (Fab.). Toxicon. 2011 Oct;58(5):415-25. doi: 10.1016/j.toxicon.2011.06.001. Epub 2011 Jul 18. PMID: 21787800.
57. Sahayaraj K, Borgio JF, Muthukumar S, Anandh GP. Antibacterial activity of *Rhynocoris marginatus* (Fab.) and *Catamirus brevipennis* (Servile) (Hemiptera:



- Reduviidae) venoms against human pathogens. *J. Venom. Anim. Toxins incl. Trop. Dis.* 2006;12:487-496. doi: 10.1590/S1678-91992006000300011
58. Bernays EA, Chapman RF, Lamunyon CW, Hartmann T. Taste receptors for pyrrolizidine alkaloids in a monophagous caterpillar. *J Chem Ecol.* 2003 Jul;29(7):1709-22. doi: 10.1023/a:1024239201198. PMID: 12921447.
59. Rousseaux CG, Schachter H. Regulatory issues concerning the safety, efficacy and quality of herbal remedies. *Birth Defects Res B Dev Reprod Toxicol.* 2003 Dec;68(6):505-10. doi: 10.1002/bdrb.10053. PMID: 14745988.
60. Rojas B, Valkonen J, Nokelainen O. Aposematism. *Current biology.* 2015;25:350-351. <https://tinyurl.com/s5b55hch>
61. Nishida R. Sequestration of defensive substances from plants by Lepidoptera. *Annu Rev Entomol.* 2002;47:57-92. doi: 10.1146/annurev.ento.47.091201.145121. PMID: 11729069.
62. Weller SJ, Jacobsen NL, Conner WE. The evolution of chemical defenses and mating systems in tiger moths (Lepidoptera: Arctiidae). *Biol J Linn Soc.* 1999;68:557-578. <https://tinyurl.com/7995ejks>
63. Pinto AF, Berger M, Reck J Jr, Terra RM, Guimarães JA. *Lonomia obliqua* venom: In vivo effects and molecular aspects associated with the hemorrhagic syndrome. *Toxicon.* 2010 Dec 15;56(7):1103-12. doi: 10.1016/j.toxicon.2010.01.013. Epub 2010 Jan 28. PMID: 20114060.
64. Kowacs PA, Cardoso J, Entres M, Novak EM, Werneck LC. Fatal intracerebral hemorrhage secondary to *Lonomia obliqua* caterpillar envenoming: case report. *Arq Neuropsiquiatr.* 2006 Dec;64(4):1030-2. doi: 10.1590/s0004-282x2006000600029. PMID: 17221019.
65. Waismam K, Chudzinski-Tavassi AM, Carrizo-Carvalho LC, Pacheco MT, Farsky SH. Lopap: a non-inflammatory and cytoprotective molecule in neutrophils and endothelial cells. *Toxicon.* 2009 May;53(6):652-9. doi: 10.1016/j.toxicon.2009.01.031. PMID: 19673080.
66. Prezoto BC, Maffei FH, Mattar L, Chudzinski-Tavassi AM, Curi PR. Antithrombotic effect of *Lonomia obliqua* caterpillar bristle extract on experimental venous thrombosis. *Braz J Med Biol Res.* 2002 Jun;35(6):703-12. doi: 10.1590/s0100-879x2002000600011. PMID: 12045836.
67. Piel J. A polyketide synthase-peptide synthetase gene cluster from an uncultured bacterial symbiont of *Paederus* beetles. *Proc Natl Acad Sci U S A.* 2002 Oct 29;99(22):14002-7. doi: 10.1073/pnas.222481399. Epub 2002 Oct 14. PMID: 12381784; PMCID: PMC137826.
68. Capineira JL. *Dermatitis linearis*. *Encyclopedia of Entomology.* Springer. 2008:1179. doi: 10.1007/978-1-4020-6359-6
69. Kellner RL, Dettner K. Allocation of pederin during lifetime of *Paederus* rove beetles (Coleoptera: Staphylinidae): Evidence for polymorphism of hemolymph toxin. *J Chem Ecol.* 1995 Nov;21(11):1719-33. doi: 10.1007/BF02033672. PMID: 24233825.
70. Kellner RL, Dettner K. Differential efficacy of toxic pederin in deterring potential arthropod predators of *Paederus* (Coleoptera: Staphylinidae) offspring. *Oecologia.* 1996 Aug;107(3):293-300. doi: 10.1007/BF00328445. PMID: 28307257.
71. Frank JH, Kanamitsu K. *Paederus*, sensu lato (Coleoptera: Staphylinidae): natural history and medical importance. *J Med Entomol.* 1987 Mar;24(2):155-91. doi: 10.1093/jmedent/24.2.155. PMID: 3295241.
72. Kellner RL. Suppression of pederin biosynthesis through antibiotic elimination of endosymbionts in *Paederus sabaeus*. *J Insect Physiol.* 2001 Apr;47(4-5):475-83. doi: 10.1016/s0022-1910(00)00140-2. PMID: 11166312.
73. Singh G, Yousuf Ali S. *Paederus dermatitis*. *Indian J Dermatol Venereol Leprol.* 2007 Jan-Feb;73(1):13-5. doi: 10.4103/0378-6323.30644. PMID: 17314440.
74. Mathes EF, Frieden IJ. Treatment of molluscum contagiosum with cantharidin: a practical approach. *Pediatr Ann.* 2010 Mar;39(3):124-8, 130. doi: 10.3928/00904481-20100223-03. PMID: 20302243.
75. Puschett JB, Agunanne E, Uddin MN. Emerging role of the bufadienolides in cardiovascular and kidney diseases. *Am J Kidney Dis.* 2010 Aug;56(2):359-70. doi: 10.1053/j.ajkd.2010.01.023. Epub 2010 Apr 22. PMID: 20417001.
76. Agunanne E, Horvat D, Harrison R, Uddin MN, Jones R, Kuehl TJ, Ghanem DA, Berghman LR, Lai X, Li J, Romo D, Puschett JB. Marinobufagenin levels in preeclamptic patients: a preliminary report. *Am J Perinatol.* 2011 Aug;28(7):509-14. doi: 10.1055/s-0031-1272965. Epub 2011 Mar 4. PMID: 21380994.
77. Eisner T, Wiemer DF, Haynes LW, Meinwald J. Lucibufagins: Defensive steroids from the fireflies *Photinus ignitus* and *P. marginellus* (Coleoptera: Lampyridae). *Proc Natl Acad Sci U S A.* 1978 Feb;75(2):905-8. doi: 10.1073/pnas.75.2.905. PMID: 16592501; PMCID: PMC411366.
78. Higashijima T, Uzu S, Nakajima T, Ross EM. Mastoparan, a peptide toxin from wasp venom, mimics receptors by activating GTP-binding regulatory proteins (G proteins). *J Biol Chem.* 1988 May 15;263(14):6491-4. PMID: 3129426.