



# International Journal of Veterinary Science & Technology

## Research article

# Serum Immunoglobulins and Lipid Profile of Sheep as Affected by Selenium and Vitamin E Supplementation - @

**Musa SI<sup>1\*</sup>, Bitto II<sup>2</sup>, Ayoade JA<sup>3</sup> and Oyedipe EO<sup>4</sup>**

<sup>1</sup>Department of Animal Science, Nasarawa State University, Keffi, Nigeria

<sup>2</sup>Department of Animal Breeding and Physiology University of Agriculture, Makurdi, Nigeria.

<sup>3</sup>Department of Animal Production, University of Agriculture Makurdi, Nigeria.

<sup>4</sup>College of veterinary medicine, University of Agriculture Makurdi, Nigeria

**\*Address for Correspondence:** Musa SI, Department of Animal Science, Faculty of Agriculture, Nasarawa State University, Keffi, Nigeria, Tel: +234-803-614-0045; E-mail: dribmazara@yahoo.com

**Submitted: 28 October 2018; Approved: 15 November 2018; Published: 20 November 2018**

**Cite this article:** Musa I, Bitto II, Ayoade JA, Oyedipe EO. Serum Immunoglobulins and Lipid Profile of Sheep as Affected by Selenium and Vitamin E Supplementation. Int J Vet Sci Technol. 2018;3(1): 026-029.

**Copyright:** © 2018 Musa I, et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



## Abstract

The effects of selenium and vitamin E administration on serum levels of immunoglobulins and lipid profile of sheep was investigated. Thirty ewes of yankasa breed were used for the study. The animals were allowed access to grazing most of the day, maize offal was provided as supplementary feed. The ewes were randomly assigned into 3 groups (n = 10). Animals in group 1 served as control and were administered 1ml normal saline. Animals in group 2 were administered 90mg Tocopherol acetate (Vitamin E), while group 3 received injection containing a combination of 100mg tocopherol acetate and 1.97mg sodium selenite. Two doses of the injections were administered 14 days apart (subcutaneously). The results indicate higher ( $p < 0.05$ ) serum values of immunoglobulin G (10.02, 11.51, 12.85) and immunoglobulin M (4.50, 5.65, 6.82) in response to a combination of selenium and vitamin E. The mean values of immunoglobulin A (1.35, 1.97, 1.70) was however similar ( $p > 0.05$ ) for all groups. Mean CD4 count values was also enhanced ( $p < 0.05$ ) following administration of a combination of selenium and vitamin E (449, 462, 498). Mean serum values of total cholesterol (3.12, 3.05, 3.00), high density lipoprotein cholesterol (1.57, 1.42, 1.67), low density lipoprotein cholesterol (1.46, 1.42, 1.1) and triglycerides (0.17, 0.27, 0.42) were similar ( $p >> 0.05$ ) for all groups. It can be concluded that supplementation with selenium and vitamin E resulted in increased serum concentration of immunoglobulin G and immunoglobulin M indicating improved immunological status of yankasa sheep. Supplementation with selenium and vitamin E can be applied in pregnant animals to improve colostrum immunoglobulin concentration which has the potential to enhance immunological status and performance of newborn animals.

**Keywords:** Immunoglobulins; Lipid profile; selenium; Vitamin E

## INTRODUCTION

Oxidative stress occurs at the cellular level, when reactive metabolites of oxygen are produced faster than they can be safely removed by antioxidant defense mechanisms. These reactive oxygen species are produced during normal metabolism, and can accumulate rapidly in actively reproducing cells. Vitamin E functions as an intra-cellular antioxidant scavenging for free reactive oxygen and lipid hydroperoxides, and converting them to non-reactive forms, thus maintaining the integrity of membrane phospholipids against oxidative damage and peroxidation. Selenium, on the other hand, functions as cofactor of the Glutathione Peroxidase \_GSH-Px. enzyme systems responsible for regulating extra and intra-cellular hydroperoxidase [1]. Oxidative stress results in macromolecular damage and is implicated in various disease states such as atherosclerosis, diabetes, cancer, neuro degeneration, and aging [2]. Protection of the body against free radicals is provided by some enzymes concerned solely with the detoxification of these radicals. Superoxide Dismutase (SOD), Glutathione Peroxidase (GPX) and catalase are the key enzymatic antioxidants of this defense system by which the free radicals that are produced during metabolic reactions are detoxified [3].

Diseases associated with selenium deficiency have been widely documented and include white muscle disease, poor reproduction, reduced daily gain and immune function. Soil Selenium concentration in the study area have been reported to be below the critical level of 0.5mg/Kg [4]. Selenium deficiency have also been reported in legumes and crop residue in the area [5]. Evidence in many species indicates that vitamin E is an essential nutrient for the normal function of the immune system [6]. Vitamin E and selenium has been implicated in stimulation of cellular and humoral immune responses [7]. Selenium deficiency plays a role in numerous economically important livestock diseases, problems that include impaired fertility, abortion, retained placenta and neonatal weakness. Administration of Se improves daily weight gain of lambs and reproductive performance in ewes [8]. Similarly reported positive influence of selenium and vitamin E on passive immune response of pregnant ewes. The adverse effect of selenium deficiency resulting in decreased IgG AND IgM had been documented [9-10]. The objective of this study is to evaluate the effects of selenium and vitamin E administration on serum levels of immunoglobulins and lipid profile of yankasa sheep.

## MATERIALS AND METHODS

### Experiment site

This study was conducted in the livestock farm of the Department of Animal Science of College Agriculture, Lafia, and Nasarawa state. The location lies within latitude 08°33' N and longitude 08°33'E at an altitude of 181.53 m (570ft) above sea level with an annual rainfall of 1311.75 cm.

### Animals and management

Thirty post pubertal Yankasa ewes (1-1½ yrs of age) weighing 22.5-26.1kg were used for the study. The animals were allowed access to grazing most of the day. Maize offal was provided as supplementary feed and Minerals salt lick and clean drinking water provided *ad libitum*. All animals were given prophylactic treatment against ecto and endo parasites, by using ivermectin (50 µg/kg subcutaneously).

### Experimental procedure

The animals were kept for a stabilization period of 14 days before commencement of the experiment. The Ewes were divided into 3 groups with 10 ewes per group. Animals in group 1 serve as control and were administered 1ml normal saline, animals in group 2 were administered injections of 90mg Tocopherol Acetate (Vitamin E), manufactured by Laborate Pharmaceutical (India). Ewes in group 3 received injections containing 100mg Tocopherol Acetate (Vitamin E) and 1.97 mg Sodium Selenite (Bremer Pharma GMBH) Germany. All injections were administered subcutaneously. Two doses of the injections was administered 14 days apart.

### Lipid profile

Triglycerides was determined using calorimetric enzymatic method with glycerophosphate oxidase, as described by [11]. Standard commercial test kits manufactured by ERBA diagnostics, Mannheim GmbH, Germany was used. Total cholesterol was determined by calorimetric enzymatic end point using reagents manufactured by Agappe diagnostics Switzerland, GmbH. HDL Cholesterol was measured by phosphotungstic acid method using commercial test kits produced by Erba diagnostics.

### Immunological parameters

Serum levels of IgG, IgM and IgA were determined using EMP-168 semi-automatic analyzer. CD4 counts was evaluated using fast count machine.

**Statistical analysis**

The data were analyzed by analysis of variance using Statistical Package for Social Science (SPSS) version 22.0. The separation of means was effected using Duncan's Multiple Range Test (DMRT) method while statement of significance were based on  $p < 0.05$ .

**RESULTS AND DISCUSSION**

Summary of effects of treatment on immunological parameters are shown on [Table 1]. Mean values of Immunoglobulin G and Immunoglobulin M are significantly higher ( $p < 0.05$ ) in group treated with selenium and vitamin E compared to the control. However, the result indicate that treatment had no effect mean values of Immunoglobulin A. Selenium and vitamin E administration resulted in significantly higher CD4 counts ( $p < 0.05$ ).

The effects of treatment on Lipid profile is shown on [Table 2]. The results indicate that administration of selenium and vitamin E had no significant effect on the serum concentration of total cholesterol. High density lipoprotein cholesterol, low density lipoprotein cholesterol and triglycerides ( $p > 0.05$ ).

Administration of vitamin E and selenium resulted in a higher ( $p < 0.05$ ) serum concentration of immunoglobulin G and immunoglobulin M, whereas, the serum level of immunoglobulin A is similar for all groups. Selenium and vitamin E administration also induced higher ( $p < 0.05$ ) levels of CD4 counts compared to that of other groups. Repeated vitamin E supplementation was reported to result in increased immunoglobulin G concentration in pregnant ewes [12]. Similarly selenium and vitamin E supplementation resulted in improved immune functions [13]. Higher serum IgG following administration of vitamin E and selenium had also been reported by [14]. This findings is very significant due to the fact that transfer of immunity from dam to neonates is hindered by placental barrier in ruminants. The lambs therefore depends entirely on antibodies obtained via colostrums. Inadequate transfer of passive immunity to lambs via colostrum has significant effect on neonatal morbidity and mortality [15-16]. Ruminant placental structure does not allow

passage of IgG from mother to fetus. The lambs are born with negligible serum concentration of IgG due to this placental barrier, they therefore depend entirely on successful transfer of colostral IgG for provision of immunity in the early days of life [9].

Mean values of total cholesterol, high density lipoprotein cholesterol, low density lipoprotein cholesterol and triglycerides were not affected by selenium and vitamin E administration. ( $p < 0.05$ ). This differ from the report of who observed a significant decrease in total cholesterol concentration and increased high density lipoprotein cholesterol in response to vitamin E and selenium supplementation in sheep [17]. This variation in response may be accounted for by the differences in duration and dosage of selenium and vitamin E administered.

**CONCLUSION**

Selenium and vitamin E are integral components of the antioxidant defense system, thereby playing significant roles in the growth and health of humans and animals. Supplementation with selenium and vitamin E had been reported to improve antioxidant status and immune functions. This study indicate that administration of combination of selenium and vitamin E resulted in increased serum concentration of immunoglobulin G, immunoglobulin M and CD4 counts indicating improved immunological status in yankasa sheep. The serum lipid profile was not affected by selenium and vitamin E.

**REFERENCES**

1. Wahiba A, Nesrine A, Fatiha B, Melhidine B, Faiza T, Amel B, et al. Modulatory role of Selenium and Vitamin E , Natural antioxidants against Bisphenol A - induced oxidative stress in Winster Albino Rats. *Toxicol Res.* 2008; 34: 231-239. <https://goo.gl/z5yGF2>
2. Gabriele P, Natasha I, Mariapoala C, Giovanni P, Federica M, Vincenzo A, et al. Oxidative stress: harms and benefits for human health. *Oxidative Medicine and cellular longevity.* 2017; 1: 13. <https://goo.gl/p2mJpG>
3. Jeeva JS, Sunitha J, Ananthalakshmi R, Rajkumari S, Ramesh M, Krishnan R. Enzymatic antioxidants and its role in oral diseases. *J Pharm Bioallied Sci.* 2015; 7: 331-333. <https://goo.gl/1zsPsu>
4. Ahola DO, Rufus S, Ishaq SE, Michael HI. Levels of Selenium in vegetables, medicinal plants and soil from selected sites within the lower Benue River Basin Catchment area, Nig. *J Environ Sci Toxic Food Technol.* 2017; 11: 21-30. <https://goo.gl/eaV8k9>
5. Adeniyi MJ, and Agoreyo FO. Nigeria and the Selenium micronutrient: A Review. *Annals of medical and health Sciences Research.* 2018; 8: 5-11. <https://goo.gl/nddzMQ>
6. Habibian M, Ghazi S, Moeini MM, Abdolmohammadi A. Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermo neutral or heat stressed conditions. *Int J Biometeorol.* 2014; 58: 741-752. <https://goo.gl/nzD5em>
7. Mohammed SA, Samar A, Emad Y, Said AZ. Effects of levamisol and vitamin E/selenium on bovine cellular and humoral immunity after bovine viral diarrhoea vaccination. *J Dairy Vet and Anim Sci.* 2015; 2: 24-28. <https://goo.gl/v6FoSc>
8. Donneml, Randy AT, Hekdoen L, Avdem F, Meling S, Vage A, et al. Effects of vitamin E supplementation to ewes in late pregnancy on the rate of stillborn lambs. *Small Ruminant Research.* 2015; 125: 154-162. <https://goo.gl/Lgrkfk>
9. Moeini MM, Jalilian MT. Effect of selenium and vitamin e injection during late pregnancy on immune system and productive performances of sanjabi ewes and their lambs, Glo. *J Anim Sci Res.* 2: 210-219.
10. Suchy P, Straova E, Herzig I. Selenium in poultry nutrition: A review. *Czech Anim Sci.* 2014; 59: 495-503. <https://goo.gl/g74hre>

**Table 1:** Effects of selenium and Vitamin E on Serum levels of Immunoglobulin of Yankasa sheep (mean + sem).

Parameters	Control	Treatment		
		Vitamin E	Se + Vit. E	
IgG(g/l)	10.02 ± 0.78 <sup>a</sup>	11.57 ± 0.47 <sup>ab</sup>	12.85 ± 0.93 <sup>b</sup>	*
IgM(g/l)	4.50 ± 0.30 <sup>a</sup>	5.65 ± 0.51 <sup>ab</sup>	6.82 ± 0.40 <sup>b</sup>	*
IgA(g/l)	1.35 ± 0.25	1.97 ± 0.25	1.70 ± 0.18	ns
CD <sub>4</sub> counts (cell / ml)	449.50 ± 44.05 <sup>a</sup>	462.50 ± 33.96 <sup>a</sup>	698.00 ± 34.36 <sup>b</sup>	*

Means within same row bearing different superscript are significantly different. SEM – Standard Error or Mean, NS = Not significant  
\* - Significant ( $P < 0.05$ ).

**Table 2:** Effects of selenium and Vitamin E on Serum lipid profile of Yankasa sheep (mean + sem).

Parameters (mmole/l)	Control	Treatment		
		Vitamin E	Se + Vit. E	
Total cholesterol	3.12 ± 0.25	3.05 ± 0.16	3.00 ± 0.18	ns
HDL-Cholesterol	1.57 ± 0.06	1.42 ± 0.13	1.67 ± 0.09	ns
LDL – cholesterol	1.46 ± 0.28	1.42 ± 0.09	1.13 ± 0.20	ns
Triglycerides	0.17 ± 0.04	0.27 ± 0.07	0.42 ± 0.16	ns

Means within same row bearing different superscript are significantly different ( $P < 0.05$ ).  
SEM – standard error of mean, ns = Not significant  
HDL – High Density Lipoprotein, LDL – Low Density Lipoprotein.

11. Manafa PO, Aquiyi NC, Onyenekwe CC, Chukwuma GO, Okeke CO, Ihim RC. Comparative assessment of lipid profile in pre -menopausal women in Nnewi, Nigeria. *European Scientific Journal*. 1:30
12. Saritha A, Christina SP, Gerald FC, Katherin HP. Effect of vitamin E in the immune system of ewes during late pregnancy and lactation. *Small Rum Res*. 2013; 111: 83-89. <https://goo.gl/1vSqe7>
13. Alhidary IA, Shini S, Aljassim RA, Abudabos AM, Gaughan JB. Effects of selenium and vitamin E on performance, physiological response and selenium balance in heat stressed sheep. *J Anim Sci*. 2013; 93: 576-588. <https://goo.gl/ybptmq>
14. Soliman EB. Dose - response of vitamin E and selenium injection on growth performance, physiological and immune responses of ossimi lambs. *Egypt J Sheep & Goat Sci*. 2015; 10: 27-40. <https://goo.gl/n3zYbC>
15. Novoa M, Aanensen L, Lind V, Larsen HJS, Jensen SK, Govasmak E, et al. Immunological effects of feeding macroalgae and various Vitamin E supplements in Norwegian white sheep ewes and their offspring. *Livestock Science*. 2014; 167: 126-136. <https://goo.gl/cHCfkz>
16. Musa SI, Bitto II, Ayoade JA and Oyedipe EO. Effects of Vitamin E and selenium on fertility and lamb performance of yankasa sheep. *Open Journal of Veterinary Medicine*. 2018; 8: 167- 173. <https://goo.gl/CW24uL>
17. Adebisi OA, Oludare OF, Majekodunmi B, Adeniji OA. Effect of vitamin e or selenium on blood profile and oxidative stability of turkey meat. *J Anim Pro Adv*. 2014; 4: 469-475. <https://goo.gl/qGf7ue>